

HIGH RESOLUTION ^1H AND ^{13}C NMR INVESTIGATIONS

OF CYCLOPHOSPHAMIDE CONFORMATIONAL MOBILITY

William Egan*

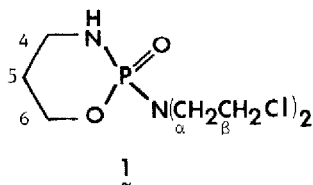
Reproduction Research Branch, National Institute of Child Health and Human Development
National Institutes of Health, Bethesda, MD 20014

Gerald Zon

Maloney Chemical Laboratory, The Catholic University of America
Washington, D C. 20064

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In connection with determining the enzymatic stereospecificity for *in vivo* activation (C-4 hydroxylation) of the anticancer drug cyclophosphamide ($\mathbf{1}$),¹ it became necessary to evaluate the dynamic conformational properties of the six-membered heterocyclic skeleton. We now report the results of high resolution ^1H and ^{13}C nmr studies with $\mathbf{1}$ which are consistent with a relatively low barrier-height for ring reversal. The present findings are in



accord with studies² of related 1,3,2-oxazaphosphorinane derivatives, but contrast several reports³ suggesting conformational rigidity in systems akin to $\mathbf{1}$.

Freshly prepared samples of $\mathbf{1}$ (chromatographed on silica gel, CHCl_3 / $\text{CH}_3\text{OH} = 9/1$) were analyzed at various temperatures by high resolution ^{13}C (68 MHz) and ^1H (220 MHz) nmr spectroscopy. As indicated in the Table, only one set of ^{13}C resonances were evident for either D_2O , CD_2Cl_2 , or CDCl_3 solutions of $\mathbf{1}$ (ca. 0.5 M) at ambient probe temperature (40°).⁴ Cooling the latter solution to -60° caused minor shifts in the resonance positions and loss of fine splitting arising from ^{31}P - ^{13}C spin-spin coupling, however, no additional signals were detected. Changing the solvent to acetone- d_6 also yielded one set of ^{13}C resonances at -60°.

Detailed investigation of the temperature dependence of the ^1H nmr spectra of $\mathbf{1}$ in various solvents was hampered by complex spin-spin coupling patterns and sample decomposition at elevated temperatures.⁵ Nevertheless, several significant conclusions regarding the con-

Table. ^{13}C NMR Parameters for Cyclophosphamide^a
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| Solvent                            | Temp. | C-4         | C-5         | C-6         | C- $\alpha$ | C- $\beta$  |
|------------------------------------|-------|-------------|-------------|-------------|-------------|-------------|
| D <sub>2</sub> O                   | 40°   | 42.90 (3.7) | 27.16 (5.6) | 71.44 (5.6) | 44.24       | 49.60 (3.7) |
| CD <sub>2</sub> Cl <sub>2</sub>    | 40°   | 41.68 (2.9) | 26.28 (6.1) | 68.28 (6.7) | 42.70       | 49.20 (4.2) |
| CDCl <sub>3</sub>                  | 40°   | 41.44 (3.1) | 25.85 (6.1) | 67.76 (7.7) | 42.32       | 48.92 (4.6) |
| CDCl <sub>3</sub>                  | -60°  | 41.24       | 25.44       | 68.13       | 41.95       | 47.29       |
| (CD <sub>3</sub> ) <sub>2</sub> CO | -60°  | 42.40       | 27.00       | 69.13       | 43.00       | 49.02       |

<sup>a</sup> Chemical shifts are in ppm downfield from internal TMS or external TSP (for D<sub>2</sub>O). Values in parentheses are  $|J_{\text{CP}}|$  in Hz, proton decoupled spectra obtained at 68 MHz. Assignments for C- $\alpha$  and C- $\beta$  are tentative

formational dynamics of  $\lambda$  derive from these <sup>1</sup>H studies. The relative signal area of each resonance multiplet allowed its assignment to either an individual hydrogen atom or group of atoms, in no instance were non-integral ratios of signal groupings observed. Additionally, when a given multiplet pattern was clearly resolved, the number of lines did not exceed that expected on the basis of a simple first-order coupling analysis. These observations provide telling evidence for the existence in solution of either a single "frozen" diastereomer, or a rapidly interconverting set of diastereomers, and are in accord with our more direct <sup>13</sup>C results. The <sup>1</sup>H nmr spectrum of  $\lambda$  was, however, temperature dependent, in that the appearance of several resonance multiplets, upon sample cooling, underwent simplification without significant resonance broadening. This was particularly evident for the  $\alpha$ -CH<sub>2</sub> protons (see Figure). While such behavior is not to be expected for the freezing out of a conformational isomer, it is explicable in terms of perturbing a distribution of conformers which are rapidly interconverting on the nmr time-scale<sup>7</sup>

A low barrier-height to ring reversal in  $\lambda$  would, by extension, be applicable to related systems and thus accommodate high resolution <sup>1</sup>H and <sup>13</sup>C data for diversely substituted cyclophosphamide derivatives, which we have found without exception to give only one set of <sup>1</sup>H and <sup>13</sup>C resonances.<sup>1,8</sup> Special substituent effects could, of course, seriously restrict conformational mobility, however, the nature of such effects are not obvious to us in cases related to  $\lambda$  where rigidity has been claimed<sup>3</sup>

Cyclophosphamide thus presents itself to the activating enzyme system as a "floppy" substrate, interpretation of data obtained in an effort to probe intimate details of the substrate-enzyme interaction must take this into account. Further insights into the conforma-

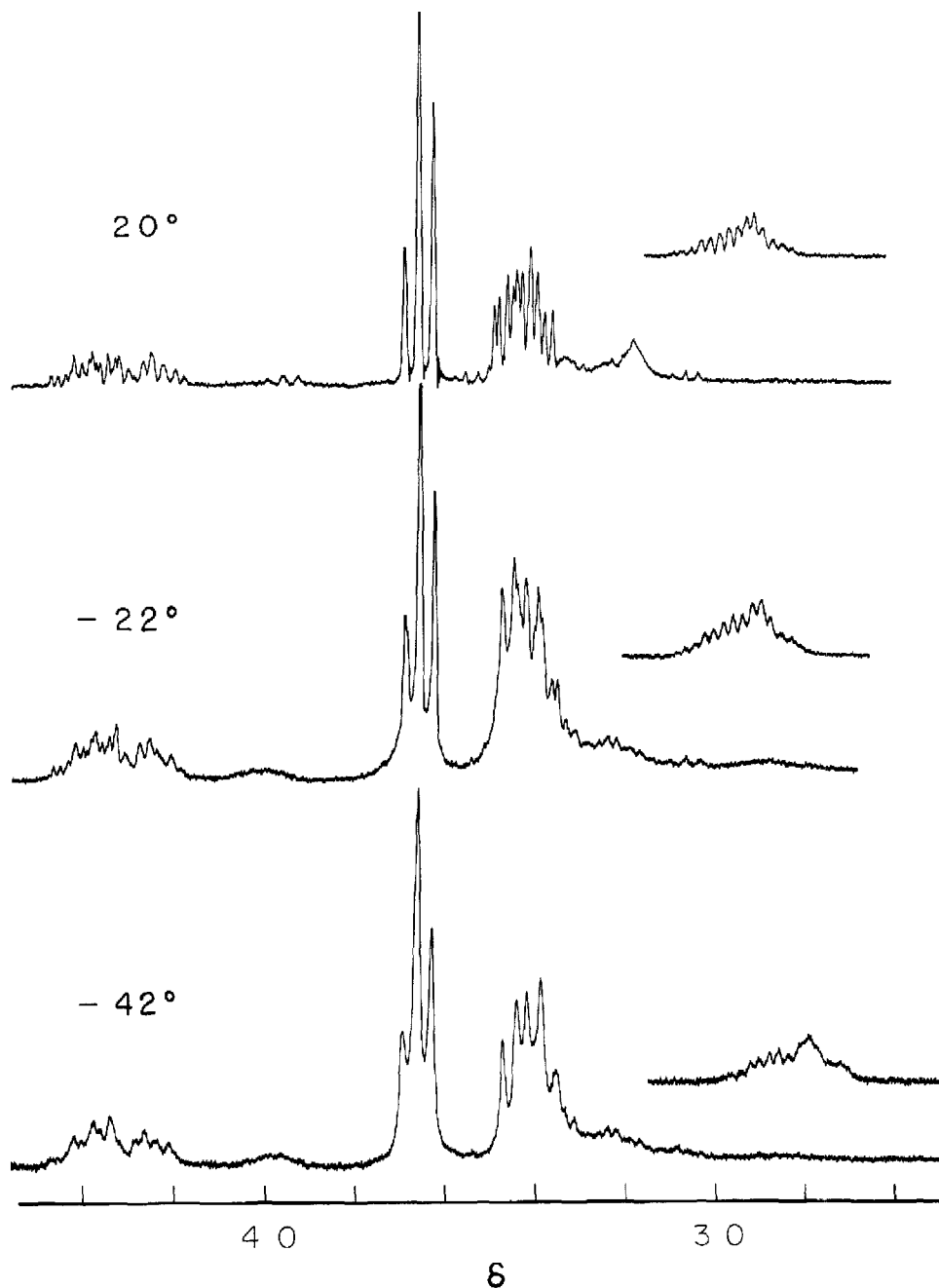


Figure. 220 MHz  $^1\text{H}$  NMR spectra of 1 in  $\text{CD}_2\text{Cl}_2$  (TMS) at various temperatures. Proton assignments:  $\delta$  4.5-4.2, C-6, 3.65 (t), C- $\beta$ , 3.5 - 3.3, C- $\alpha$ , 3.3 - 3.1, C-4, 1.85 - 1.95 (offset), C-5, N-H variable

tional behavior of  $\downarrow$  are being pursued by means of molecular orbital calculations<sup>9</sup>

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